

A Computer Model
of the Cellular Slime Mould
Dictyostelium discoideum

Roger P. Stevens, BE (Mech.)
School of Computing & Information Technology
Griffith University

Submitted in fulfilment of the requirements of the degree of
Master of Philosophy

9 September, 2002

Statement of Originality

This work has not previously been submitted for a degree or diploma in any university. To the best of my knowledge and belief, the thesis contains no material previously published or written by another person except where due reference is made in the thesis itself.

Signed:

Acknowledgements

I wish to acknowledge all those who assisted with the prosecution of this thesis, including my wife who suggested I undertake the degree, the staff at QPSF, my work supervisor, but particularly my academic supervisors.

Abstract

Excitable media are an important class of systems, examples of which include epidemics, predator-prey interactions, nervous systems, and heart muscle. Aggregating cellular slime moulds are an example of an excitable medium. The species of cellular slime mould *Dictyostelium discoideum* is an important model organism that many science laboratories use. Studying the aggregation of slime moulds increases knowledge about excitable media generally. One method of studying the aggregation of slime mould is to simulate their behaviour on a computer model.

This thesis presents the author's computer model of cellular slime mould *Dictyostelium discoideum* and the results of experiments carried out using the computer model. The experiments investigate the relation between the aggregation patterns and the various parameters of the model. These parameters are the density of artificial slime moulds, the acrasin threshold, the acrasin degradation rate, and the rate of acrasin secretion. Randomness has an effect on the aggregation patterns produced. Results of experiments are presented that examine the effect of randomness. Two forms of randomness are investigated: random secretion of acrasin by the artificial slime moulds; random initial reactivity of the artificial slime moulds.

The computer model describes an artificial environment in which artificial slime mould amoebae interact with each other and their environment. Out of these individual interactions the global patterns that characterize slime mould aggregations emerge. The model facilitates the study of these individual interactions and hence the global patterns that emerge.

The model and the experimental results described in this thesis contribute to the study of the aggregation phase of the life cycle of *Dictyostelium discoideum*. The author proposes mechanism that could underlie certain classes of aggregation patterns. These patterns include net-like aggregations and loop aggregations.

The computer model presented in this thesis is successful in emulating the behaviour of the cellular slime mould *Dictyostelium discoideum*. In its present form the model is a useful tool to biologists. The results of experiments conducted with the model suggest mechanisms that may underlie certain pattern produced by living slime moulds. A result of particular interest is the initiation of the spiral wave pattern from a loop wave, which produces a loop aggregation.

Table of Contents

Statement of Originality.....	I
Acknowledgements	II
Abstract.....	III
Table of Contents	IV
List of Figures	VII
List of Tables	IX
Notation	X
1.0 Introduction.....	1
1.1 Background	1
1.2 Aims	2
1.3 Significance.....	2
1.4 Key Terms.....	3
1.5 Organisation of Thesis.....	3
2.0 Literature Review.....	4
2.1 Introduction	4
2.2 Parameters Determining Acrasin Wave Propagation and Slime Mould Aggregation.....	5
2.3 Acrasin Wave Propagation Patterns	7
2.3.1 Concentric Circular Waves.....	7
2.3.2 Spiral Waves.....	7
2.3.3 Loop Waves.....	8
2.4 Slime Mould Aggregation Patterns	8

3.0	Design and Implementation	15
3.1	Introduction	15
3.2	Overview of Model	16
3.3	Overview of a Simulation.....	18
3.4	Modelling of the Environment	19
3.4.1	Chemical Diffusion.....	20
3.4.2	Chemical Degradation	22
3.5	Modelling of Amoebae.....	22
3.5.1	Stage of Life Cycle	23
3.5.2	Energy of an SM.....	24
3.5.3	CAMP Receptors	24
3.5.4	cAMP Relaying.....	26
3.5.5	Speed of Movement.....	26
3.5.6	Direction of Movement.....	27
3.5.7	SM Location	29
3.5.8	Time Since the Last cAMP Relay Event.....	29
3.5.9	The Number of cAMP Relay Events	30
3.6	Modelling of Bacteria.....	30
4.0	Experiments, Results and Discussion	31
4.1	Model Calibration.....	31
4.1.1	Diffusion	31
4.1.2	Wave Speed and Continuity.....	35
4.2	Input Parameters.....	41
4.3	Output.....	45
4.3.1	Field Visualizations Output	45
4.3.2	cAMP Wave Cross Sections Output	47
4.3.3	Wave Speed and Continuity Output.....	48
4.4	Experiments.....	48
4.4.1	Aggregations to a Pacemaker SM.....	48
4.4.2	Random Secretion of cAMP	62
4.4.3	Formation of Natural Loop Aggregation	63
4.4.4	Loop Dynamics Experiments.....	66
5.0	Further Research	69
6.0	Conclusion.....	71

References.....	73
Appendix 1: Discrete Diffusion	80
Appendix 2: Calculating Maximum Gradient	82
Appendix 3: Slime Mould Parameter Values.....	84
Appendix 4: Glossary.....	86
Appendix 5: Cellular Slime Mould Biology	90
Classification	90
Life History.....	91
Feeding	92
Aggregation	93
Pseudoplasmodium	96
Culmination	97
Sori.....	97
Microcysts.....	98
Sex and Macrocyts	98
Adaptive Advantage of Life Cycle	98
Ecology.....	99
Appendix 6: Notes of Electronic Media.....	100
List of Files on Windows CD	100
List of Files on Macintosh CD.....	100
Field Visualisation Application	100

List of Figures

Figure 2.1 A segment of an aggregation with streams with little branching	9
Figure 2.2 A portion of an aggregation with branching streams	10
Figure 2.3 A segment of an aggregation where streams merge towards the centre.....	11
Figure 2.4 An aggregations with interconnecting and detached streams.....	12
Figure 2.5 A loop aggregation.....	13
Figure 2.6 A network of interconnecting streams without a defined centre	13
Figure 3.1 Setup of acrasin wave cross-section experiments	17
Figure 3.2 Setup of experiments to measure the speed and continuity of acrasin wave.....	18
Figure 3.3 The field is an artificial two dimensional surface on which SM 'live'.....	20
Figure 3.4 Calculation of diffusion.....	21
Figure 3.5 The relationship between cAMP concentration and the reactivity of a SM plotted against time	25
Figure 3.6 An SM about to move	26
Figure 3.7 The direction of maximum concentration gradient	28
Figure 3.8 The orientation of an SM	29
Figure 4.1 Diffusion with no degradation from a single central field cell.....	32
Figure 4.2 Modelled diffusion for two values of r_{xy}	33
Figure 4.3 Comparing diffusion from the model with a calculated distribution from a point source....	34
Figure 4.4 cAMP wave speed, cAMP wave speed standard deviation, & number of triggered wave check field cells plotted against the number of SM2	36
Figure 4.5 cAMP wave at higher SM2 densities	37
Figure 4.6 cAMP wave at lower SM2 densities	38
Figure 4.7 Average wave speed for 4, 8, and 16 wave check field cells	39
Figure 4.8 Standard deviation of wave speed for 4, 8, and 16 wave check field cells.....	40
Figure 4.9 A cross section through the cAMP wave shown in Figure 4.5.....	47
Figure 4.10 The relation between cAMP wave speed and SM density	49
Figure 4.11 The mechanism of aggregation stream formation.....	50
Figure 4.12 The first cAMP wave at higher cAMP loss rate.....	51
Figure 4.13 The first cAMP wave at lower cAMP loss rate.....	51

Figure 4.14 The first cAMP wave at higher cAMP threshold	52
Figure 4.15 The first cAMP wave at lower cAMP threshold	52
Figure 4.16 Clump aggregations	54
Figure 4.17 Aggregation with very low cAMP degradation.....	55
Figure 4.18 Aggregation patterns for cAMP threshold of 0.000001 and various cAMP degradation rates.....	56
Figure 4.19 Aggregation patterns for cAMP threshold of 0.00001 and various cAMP degradation rates	58
Figure 4.20 Aggregation patterns for cAMP threshold of 0.0001 and various cAMP degradation rates	59
Figure 4.21 Aggregation with cAMP secretion reduced to 5 SMU.....	60
Figure 4.22 Aggregation with high cAMP degradation, high cAMP threshold concentration, and very low cAMP secretion rate.....	61
Figure 4.23 Random secretion of cAMP with one SM3.....	62
Figure 4.24 Net-like aggregation steams produced by random secretion of cAMP but with no SM3 ..	63
Figure 4.25 The beginnings of a loop aggregation	64
Figure 4.26 A loop aggregation formed by initial randomly assigned reactivity and some random firing of SM	65
Figure 4.27 Detail of a cAMP wave approaching the corner of a square loop.....	67
Figure 4.28 Detail of how the wave interacts with SM arranged in a circle.....	68
Appendix 2 Figure 1 Vector addition to obtain the vector ∇C with a direction given by β	83
Appendix 5 Figure 1 Life cycle of the cellular slime mould <i>Dictyostelium discoideum</i>	92
Appendix 5 Figure 2 Feeding amoeba.....	93
Appendix 5 Figure 3 Loop aggregation.....	96
Appendix 5 Figure 4 Mature spore.....	98

List of Tables

Table 4.1	Parameter values supplied by the ParamFileSM5	41
Table 4.2	Field visualization output modes	46
Table 4.3	Colour codes for SM reactivity	46

Notation

C	concentration of a chemical
C_a	concentration of acrasin
$C_{\text{threshold}}$	threshold concentration of acrasin above which an SM will react
D	diffusion constant
D_a	diffusion constant for the acrasin
d	diameter of an SM
n	the number of triggered wave check field cells
r	a measure of the sensitivity of SM to acrasin, i.e. corresponds to the number of unbound acrasin receptors in living slime mould amoebae, and there for the SM's reactivity
s	the speed of the cAMP wave
t	time variable
Δt_{iter}	the increment in time for each iteration of the model
v	speed of SM
v_{max}	maximum speed of SM
α	the angle
β	the angle of maximum chemical gradient
δ	distance between two SM
δ_{fc}	distance between field cells, ie the length of the side
δ_{field}	the distance from the central SM3 to the wave check field cell
ε	a small random angle added to the direction of maximum rate of change in chemical concentration with respect to distance to give the direction of movement of a SM
ϕ	the angle from direction zero to the direction of maximum rate of change in chemical concentration with respect to distance
σ	the distance a SM moves